

**VIABILITY OF WOOD DECAYING FUNGAL MYCELIUM AFTER
MICROWAVE RADIATION OF BAMBOO CULM**

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ABSTRACT

The present study was carried out to evaluate the effects of microwave (MW) radiation on viability of wood decaying fungi. The white rot (*Trametes versicolor*) and brown rot (*Rhodonia placenta*) fungi were grown on bamboo culm-samples. The mycelium growths were observed in controlled as well as microwave treated samples. The results showed that the viability of fungi decreased according to the applied MW time. This study proved the ability of the microwaves and exposure time MW3 (180 seconds) to kill the fungal colonies and do not allow for the growth of fungal spores, means the rate of growth of fungal colonies is inversely proportional to time of microwave exposure.

Keywords: Bamboo culm wood, *Bambusa vulgaris*, decay, fungi, microwave treatment, viability.

1 INTRODUCTION

2 The chief economic product of forest is wood i.e., utilized for various purposes such as
3 construction, furniture, door, window etc. Failure of wood in service through biodeterioration
4 is a prime cause of user dissatisfaction. Although many types of organisms deteriorate wood,
5 the greatest damage is from fungi and insect, and to a minor extent- from bacteria (Highley *et*
6 *al.* 1994; Schmidt 2006).

7 The most important and potent wood-decay fungi are white and brown-rot. For decay to
8 occur, the minimum moisture content is slightly below the fibre saturation range (average 25-
9 30 percent) and the temperature between about 2 °C and 45 °C (Pasanen *et al.* 1992; Highley
10 *et al.* 1994; Morris and Winandy 2002; Schmidt 2006). Stienen *et al.* (2014) showed mycelia
11 growth on wood with 17,4 % moisture content and mass loss over 2 % at 24,6 % moisture
12 content. Oxygen must also be available because decay fungi are aerobic organisms. Finally, a
13 suitable substrate, such as wood, must be present to nourish the fungus. Interference with any
14 of these requirements stops the decay process.

15 Bamboo has a low natural durability (Liese and Kumar 2003). As a biological material
16 like wood, it is susceptible to degrade by different organism such as insect and fungi
17 (Schmidt *et al.* 2011). Bamboo deterioration can be controlled by impregnation with
18 chemicals, but due to toxic behaviours of these chemicals, namely, pentachlorophenol,
19 copper chrome arsenate, creosote and several others many countries have banned their use for
20 wood preservation (Poonia and Tripathi 2016). Public concern about the use of synthetic
21 chemicals concomitantly with significantly tightened environmental regulations have created
22 demands for the development of alternative methods for the chemical wood protection that
23 we can go for substitute like microwave treatment.

24 Microwave (MW) radiation is an innovative method for improving wood permeability,
25 treatability and the drying rate of wood. This method can also be capable of inactivating

1 microbial contaminations not only on exposed surfaces but inside them as well (Poonia and
2 Tripathi 2018). It can result in reducing or stopping colonization of microbiologically
3 contaminated surfaces and thus can decrease the number of agents contributing to the adverse
4 effects on growing environment of fungi (Chipley 1980).

5 The majority of the experiments i.e., effects of microwave radiation were conducted on
6 bacteria and yeasts. There are several studies however some studies on basidiomycetes. For
7 example, Baumann-Ebert and Körner (2013) used portable MW devices to kill the most
8 important European indoor rot fungus *Serpula lacrymans* (brown rot) within thick wooden
9 beams. Plarre *et al.* (2013) performed corresponding experiments with *S. lacrymans* and
10 larvae of the beetles *Anobium punctatum* and *Hylotrupes bajulus*. There are already national
11 standards for the use of MW treatment against wood-destroying organisms. For example, the
12 German standard DIN 68800-4 (DIN 2012) allows the use of MW treatment against insects in
13 buildings, however not against fungi due to less research results.

14 The present work was carried out to evaluate the effect of MW on viability and decay of
15 future wood decaying fungi.

16

17 **MATERIALS AND METHODS**

18 **Preparation of bamboo culm specimens**

19 Bamboo (*Bambusa vulgaris* Schrad.) culms were procured from Sirsi (Lat.14° 61' N,
20 Long.74° 85' E), Karnataka, India and converted into small clear specimens of size 2 cm × 2
21 cm. The defect free specimens were randomly selected and further smoothen by sand paper
22 to remove the silica layer.

23 **Preparation of Culture media**

24 A nutrient medium (i.e., PDA) consists of 4 g/l potato starch, 20 g/l glucose and 15 g/l
25 agar was prepared in an Erlenmeyer bottle. The Erlenmeyer was then plugged with cotton,
26 wrapped with foil and autoclaved at 2,1 bar and 121 °C for 20 minutes.

1 **Inoculation of fungi**

2 The effect of microwave radiation was tested against the white rot fungus (*Trametes*
3 *versicolor* (L.: Fr.) Pilat) and the brown rot species *Rhodonia placenta* (Fr.) Niemelä, K.H.
4 Larss. & Schigel. The fungi were inoculated on potato dextrose agar (PDA) in Petri dishes
5 under a laminar air flow. Inoculated plates were then kept at $25\text{ }^{\circ}\text{C} \pm 2\text{ }^{\circ}\text{C}$ and $75\text{ \%} \pm 5\text{ \%}$
6 RH for 7 days for growth. After the growths of mycelial, spore suspension was prepared
7 according to IS: 4873 (2008).

8 Bamboo culm samples were exposed to fungal attack by placing them in the petri
9 dishes in which there are actively growing cultures of *T. versicolor* and *R. placenta*. The
10 spore suspension was sprayed over the bamboo samples for aerial contact with fungal
11 mycelium. Then, the petri dishes were further kept at $25\text{ }^{\circ}\text{C} \pm 2\text{ }^{\circ}\text{C}$ and $75\text{ \%} \pm 5\text{ \%}$ RH for
12 growth of fungi on bamboo samples. After initiation of growth, the petri dishes were kept in
13 MW oven for different times.

14 **Microwave treatment**

15 MW oven with maximum output power 900 W at frequency 2450 MHz was used.
16 Treatments were done at different times i.e., 60 s (T-1), 120 s (T-2) and 180 s (T-3). Six
17 replicates in each treatment. The controls were maintained simultaneously.

18 After MW treatment, inoculated bamboo samples were further kept at $25\text{ }^{\circ}\text{C} \pm 2\text{ }^{\circ}\text{C}$
19 and $70\text{ \%} \pm 4\text{ \%}$ RH. The results were recorded after 21 days in terms of per cent surface
20 covered by the test fungi over the bamboo samples (Tripathi *et al.* 2009). The percent
21 inhibition in growth of fungi was calculated by using visual scores and by weight loss
22 (Poonia *et al.* 2015).

23 **Weight loss and visual scores assessment of fungal growth**

24 The bamboo blocks were weighed before (W1) and after 21 days of inoculation of
25 fungi (W2). In MW treated specimens the weight after MW treatment were considered as

1 initial weight. The efficacy of treatment was categories on basis of weight loss and surface
2 coverage by test fungi. Weight loss was calculated as follows (Eq. 1):

3
$$\text{Weight loss method (\%)} = \frac{W_1 - W_2}{W_1} \times 100 \quad (1)$$

4 Where, W1= Weight of Petri dish containing bamboo blocks before inoculation of fungi (g)

5 W2= Weight of Petri dish containing bamboo blocks after 21days inoculation of fungi (g)

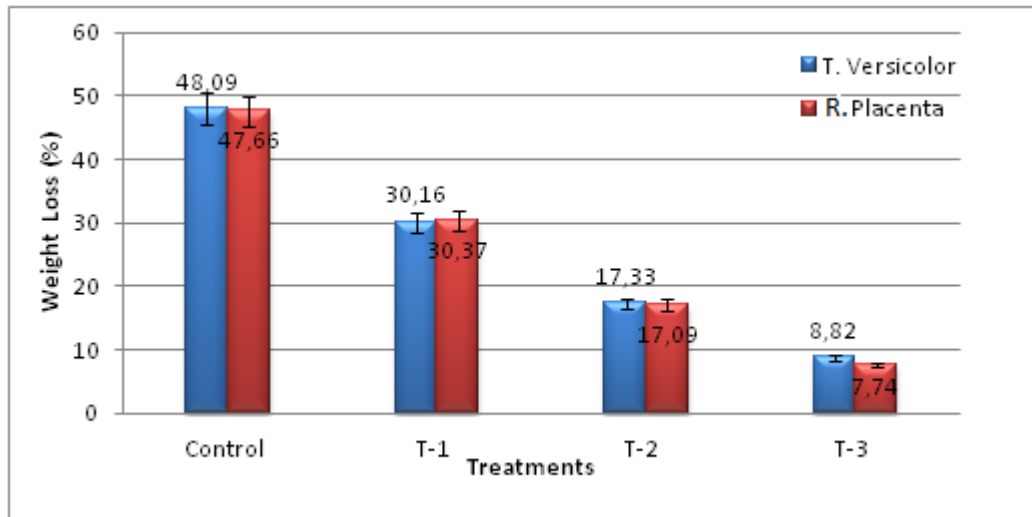
6 **Statistical analysis**

7 Statistical analysis was conducted using SPSS 16 version (IBM 2007). Effect of MW
8 treatment time on fungi growth/viability were compared using variance analysis (ANOVA)
9 and the Duncan homogeneity test at 95 % confidence interval.

10

11 **RESULTS AND DISCUSSIONS**

12 The results revealed that the control samples exhibited higher weight loss i.e., 48,09
13 and 47,66 percentage in white (*T. versicolor*) and brown rot (*R. placenta*) fungi respectively.
14 Further, it was observed that the 60 s showed less effect on growth with weight loss of 30,16
15 and 30,37 % (*T. versicolor* and *R. placenta*, respectively) as shown in Fig. 1. Whereas, the
16 weight loss percentage were significantly decreased as increased in MW treatment duration in
17 both the fungi at ($P \leq 0,05$) level. The MW treatment time with specific frequency plays an
18 important role in the effect of the microwaves which is time-dependent, where the intensity
19 of the weight loss carried by fungal infection is inversely proportional to the total absorbed
20 microwave energy.



1

2 **Figure 1:** Effect of microwave treatment on weight loss of Bamboo blocks/samples.

3 The results in term of surface coverage by test fungi i.e., *T. versicolor* and *R. placenta*
 4 over bamboo samples containing PDA medium are shown in Table 1 and Fig. 2.

5 **Table 1:** Effect of microwave treatment on bamboo sample surface coverage of wood
 6 decaying fungi.

Sample No.	Treatment	Surface coverage (%)		Efficacy of treatment
		<i>T.versicolor</i> (Tv)	<i>R.placenta</i> (Rp)	
1	Control	100 ^a	100 ^a	Growth complete (> 75 %)
2	T-1	32,50 ^b	40,50 ^b	Growth moderate (25 % to 50 %)
3	T-2	10,70 ^c	8,75 ^c	Growth Little (5 % to 25 %)
4	T-3	0,00 ^d	0,00 ^d	Growth none (0 %)

7 Mean square error of Tv and Rp is 6,34 and 6,74 respectively at ($P \leq 0,05$) level. Different
 8 letters denote significantly different groups

9 It was observed that T-1 treatment (60 s of exposure) showed less effect on growth of both
 10 fungi, the inhibition was 67,50 and 59,50 % (*T. versicolor* and *R. placenta*) respectively.

11 While the results shown that after T-3 treatment (180 s of exposure) the fungal growth were
 12 completely inhibited and the percentage of inhibition were 100 % for both fungi as compare
 13 to controls. The viability of mycelium of both fungi were significantly decreased ($P \leq 0,05$)
 14 after MW exposure. The reason behind inhibition of fungal growth may be due to heat

1 generated by microwaves create steam pressure within the wood/micro-organism due to
2 presence of moisture exerted the killing effect on fungi (Gorny *et al.* 2007).
3 The microwaves treatment at frequency 2450 MHz, with high energy and for a sufficiently
4 long period of time, their thermal effect is most likely dominant to kill fungal mycelium (Al-
5 Mayah and Ali 2010). However, at lower duration treatment, the killing effect of microwave
6 radiation was significantly decreased and happened only after a prolonged period of
7 irradiation, because of a lower transformation of microwave energy to heat. Some other
8 studies showed that the extent of killing of microorganisms was correlated with the moisture
9 content of the experimental specimens. In contrast, when microorganisms were irradiated
10 with microwaves at temperatures lower than the thermal destruction level; various effects
11 were observed, from killing to enhanced growth (Kuchma *et al.* 1992; Ahmed *et al.* 2015).
12 Hence, the study concluded that microwave radiation method has better possibilities of
13 application using wood of building constructions (Gorny *et al.* 2007). Covering a larger space
14 with microwave field create the possibility to obtain a more homogeneous temperature
15 distribution on the wood subjected to reduce the growth of fungus. So that using of
16 microwave radiation treatment we can reduce the wood deterioration.

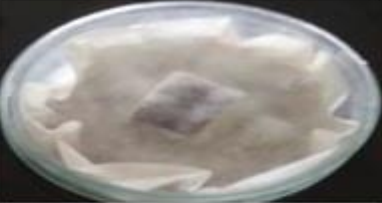
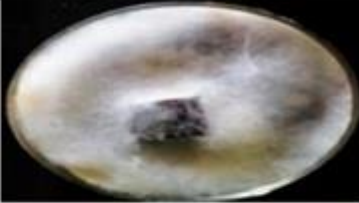
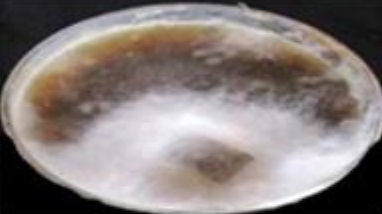

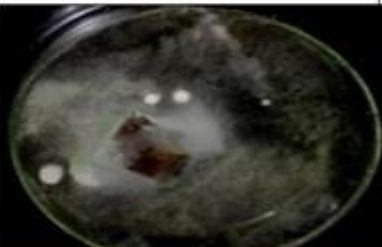


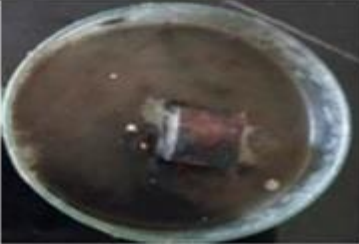
S. NO	Treatment	Surface coverage (%)	
		<i>T.versicolor</i> (Tv)	<i>R. placenta</i> (Rp)
1	Control		
2	T-1		
3	T-2		
4	T-3		

Figure 2: Surface coverage by wood decaying fungi after MW treatment.

However, it should be considered that surface coverage of wood or bamboo culm samples are no sure indicator for degradation of the respective wood tissues. There are several wood-decay fungi which produce severe wood degradation with no or only meager surface mycelium, for example, the brown cellar fungus *Coniophora puteana* in buildings (Schmidt 2006).

CONCLUSIONS

The present study was carried out to evaluate the viability of wood decaying fungi growing on bamboo culm samples against microwave treatment. The study revealed that the

1 microwave play a significant role in controlling the fungal growth/viability. Hence, this study
2 suggests that to apply this technology at pilot scale can control fungal growth in building
3 materials.

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