DOI: 10.4067/s0718-221x2021000100428

PHENOLICS CONTENT AND ANTIOXIDANT ACTIVITY OF WOOD EXTRACTIVES FROM THREE CLONES OF ACACIA HYBRID (Acacia mangium × Acacia auriculiformis)

Brandon Aristo Verick Purba¹

https://orcid.org/0000-0003-1918-2927

Sri Sunarti²

https://orcid.org/0000-0003-2968-8278

Ganis Lukmandaru^{1,}*

https://orcid.org/0000-0003-2203-4247

ABSTRACT

Breeding strategy of Acacia hybrid is being developed by Center for Forest Biotechnology and Tree Improvement Indonesia and has produced three superior clones in growth (Clone 16, 25, and 44). Understanding the extractives of the new clones might determine future development steps to improve its resistance to diseases especially heartrot. The objective of this study was to investigate the extractive content in three radial directions (SW = sapwood; OHW = outer heartwood; IHW = inner heartwood); total phenolic, flavonoid, flavanol contents (colorimetric assay); and antioxidant activity (1,1-diphenyl-2-picryl-hydrazyl (DPPH) radical scavenging assay). Extractions were done with three different solvents in successive (n-hexane = H; methanol = M; hot water = W), yielded 0.69 % - 1.70 %; 1.51 % - 10.86 %; and 0.51 % - 1.16 % of extractive contents, respectively. The total phenolic content (TPC) from TPC-H, TPC-M, and TPC-W ranged between 3,68 mg of gallic acid equivalent (GAE)/g - 10,41 mg GAE/g; 76,83 mg GAE/g - 448,35 mg GAE/g; and 43,28 mg GAE/g – 198.92 mg GAE/g, respectively; the total flavonoid content (TFC) from TFC-H, TFC-M, and TFC-W between 4,23 mg of quercetin equivalent (QE)/g - 41,51 mg QE/g; 29,55 mg QE/g - 133,71 mg QE/g; and 7,70 mg QE/g – 29,37 mg QE/g, respectively; total flavanol content (TVC) from TVC-H, TVC-M, and TVC-W ranged between 28,74 mg of catechin equivalent (CE)/g - 66,90 mg CE/g; 83,39 mg CE/g - 247,18mg CE/g; and 7,08 mg CE/g – 29,21 mg CE/g, respectively. Furthermore, the antioxidant activity was found to be significantly affected by the radial factor with the strongest activity exhibited by inner heartwood extract with an IC_{50} value of 255,77 µg/ml (gallic acid IC_{50} showed a value of 39,00 µg/ml). Among clones, clone 16 was determined to have the highest extractive, total flavonoid as well as flavanol contents. Thus, clone 16 was hypothesized to be more resistance against heart rot disease.

Keywords: Acacia hybrid, heartwood, phenolic contents, sapwood, wood extractives.

INTRODUCTION

Acacia mangium is a multipurpose fast-growing species being used in plantations throughout some tropical countries (Sein and Mitlohner 2011). Over the last few years, its productivity has decreased due to fungal attack which causes heart and root rots (Mohammed *et al.* 2006, Coetzee *et al.* 2011). Heart rot is found in the heartwood of a tree and despite the fact it does not kill it, the merchantable wood volume is reduced by up to 24 % (Sudin *et al.* 1993). However, the less preferable species of Acacia genus to be planted in production forest

¹ Universitas Gadjah Mada, Faculty of Forestry, Department of Forest Products Technology, Bulaksumur, Yogyakarta, Indonesia.

²Center for Forest Biotechnology and Tree Improvement, Purwobinangun, Pakem, Sleman, Yogyakarta, Indonesia.

^{*}Corresponding author: glukmandaru@ugm.ac.id

Received: 19.07.2019 Accepted: 19.01.2021

is Acacia auriculiformis and it is known to have more resistance to heart rot (Barry et al. 2005).

Acacia hybrid is a crossbreed, either naturally or artificially, between Acacia mangium and Acacia auriculiformis which inherits several advantages from both of its parent, including faster growth, better stem form, and better resistance against pest and disease compared to its parent species (Pinso and Nasi 1991, Kha 2000, Sunarti et al. 2013). The Center for Forest Biotechnology and Tree Improvement Indonesia has been developing a breeding strategy for Acacia hybrid using the co-improvement method as one of the efforts to increase the productivity of production forest by hybridization. This was conducted using selected plus trees of A. mangium and A. auriculiformis parent trees from different family and provenance. By the co-improvement method, it produced a total of tested 44 clones, from which three clones, 44, 16, and 25, observed to have superior growth performance were selected (Sunarti et al. 2013).

Previous studies have identified some phenolic compounds in *A. mangium* and *A. auriculiformis* wood extractive and 3,4,7,8-tetra-hydroxy flavanone and teracacidin are the most abundant flavonoid compounds found in both species to have shown antifungal activity (Pietarinen *et al.* 2004, Barry *et al.* 2005). Furthermore, the ability of these compounds to resist heart rot was attributed to the scavenging of hydroxyl radical produced by the fungi, as correlations between antifungal activity, laccase inhibition, and antioxidant activity (Mihara *et al.* 2005).

However, the information on *Acacia* hybrid wood extractive has been found to be limited to its general extractive and lipophilic content (Yahya *et al.* 2010, Soon and Chiang 2012). Therefore, in this study, the extractive content of *Acacia* hybrid clones was measured in its radial direction for total phenolic, flavonoid, and flavanol content as well as the antioxidant activity. This information can be used to indicate each *Acacia* hybrid clone quality in many aspects regarding its extractive content, including resistance to heart rot disease and as a potential source of natural antioxidant.

MATERIALS AND METHODS

Wood material

Three six-year-old trees of each *Acacia* hybrid superior clones 16, 25, and 44 were harvested from Wonogiri, Central Java, Indonesia. The trees' parent family, provenance, and clone growth data are described in Table 1. Moreover, the disc was cut from a height of 15 cm above ground level and drilled in sapwood (\pm 1 cm from the border of sapwood and bark), outer heartwood (\pm 0,5 cm from the border of sapwood and heartwood), and inner heartwood (\pm 5 cm radius of the pith) and the rough residues were collected, milled, and sieved using a 40-sized mesh.

Clone	Average breast-height diameter (cm)	Average heartwood proportion (%)	Average total height (m)	Family and Provenance (A. mangium)	Family and Provenance (A. auriculiformis)	
16	23,50	67,23	16,30	86 Claudi River, Iron 107 RA, Australia	107 Orchard Melville, Int, Queensland	
25	19,25	63,45	19,48	114 Claudi River, Iron 112 RA, Australia	112 Orchard Melville, Int, Queensland	
44	30,25	59,47	20,30	86 Claudi River, Iron 101 RA, Australia	101 Kennedy River, Queensland	

Table 1	1: Descri	otion of	each c	lone s	amples.
---------	-----------	----------	--------	--------	---------

Successive extraction

Furthermore, 5 g O.D. (oven dried) wood powder samples from each clone and radial section were extracted successively through the use of n-hexane (Soxhlet, 6 h), methanol (Soxhlet, 6 h) and hot water (water

bath, 3 h) solvents. The extractive content from each of them was calculated based on the percentage of dry weight after drying with a rotary evaporator.

Total phenolic content

The total phenolic content was calculated using Folin-Ciolcateu method according to the procedure described by Baba and Malik (2015). This was conducted by adding 0,5 ml of the extract in 1 mg/ml concentration to 2,5 ml Folin-Ciolcateu reagent (Merck, Germany) that has been diluted 10 times. After 2 min, 2 ml of 7,5 % Na₂CO₃ was also added and the solution was incubated for 30 min under room temperature and analyzed using UV-VIS spectrophotometer in 765 nm wavelength. Moreover, a calibration curve was also prepared using the same procedure with gallic acid as the standard compound and total phenols were calculated as gallic acid equivalents (GAE) and expressed as mg GAE/g sample.

Total flavonoid content

The total flavonoid content was calculated using aluminum chloride (AlCl₃.H₂O) according to the procedure described by Diouf *et al.* (2009). This was conducted by reacting a diluted 2 ml of the extract to achieve 1 mg/ml concentration with 2 ml of 2 % AlCl₃.H₂O. The solution was shaken and incubated for 30 min under 20 °C and the absorbance was measured using UV-VIS spectrophotometer in 415 nm wavelength. Moreover, a calibration curve was also prepared using the same procedure with quercetin as the standard compound and the total flavonoids were calculated as quercetin equivalents (QAE) and expressed as mg QAE/g sample.

Total flavanol content

The total flavanol content was calculated using vanillin - H_2SO_4 according to the procedure described by Diouf *et al.* (2009). This was conducted by reacting 1 ml of the extract in 1 mg/ml concentration with 2 ml of vanillin - H_2SO_4 reagent (1 g vanillin in 100 ml of 70 % H_2SO_4). The solution was incubated for 15 min under 20 °C temperature and stopped by cooling the solution using a block of ice and the absorbance was measured using UV-VIS spectrophotometer in 500 nm wavelength. Moreover, a calibration curve was also prepared using the same procedure with catechin as the standard compound and the total flavanols were calculated as catechin equivalents (CAE) and expressed as mg CAE/g sample.

DPPH radical scavenging assay

As much as 0,1 ml of methanol extract in varying concentrations of 100, 150, 200, and 250 μ g/ml was added to the solution of 3 ml of 0,1 mM 1,1-diphenyl-2-picrylhydrazyl (DPPH) (Sigma-Aldrich, USA) in methanol. The solution was shaken and kept in dark for 30 min under 22 °C temperature. Moreover, the blank solution also reacted without the addition of extract with the same procedure and the absorbance was measured at 512 nm. The calculation of inhibition was conducted using the following Equation 1:

Inhibition
$$(\%) = 100 \times \frac{(A0 - A1)}{A0}$$
 (1)

Where A0 is the absorbance of the blank and A1 is the absorbance of the sample with the extract. Antioxidant activity expressed as IC_{50} (concentration inhibiting the DPPH reaction by 50 %), however, lower IC_{50} indicates higher antioxidant activity. In this experiment, gallic acid and catechin were used as a positive control and all analyses were run in triplicate and averaged.

Statistical analysis

The data were analyzed using the SPSS program (Ver. 25, IBM). A two-way analysis of variance (ANOVA) was conducted with a 95 % confidence level to determine the effect of clone (44, 25, and 16) and radial (sapwood, outer heartwood, inner heartwood) direction on the content of extractive, phenolic, flavonoid, and flavanol, as well as the antioxidant activity of the methanol-soluble fractions. All the data were assayed as a normal data distribution. Furthermore, Duncan test was performed to evaluate which factor affected significantly while Pearson's correlation was used to measure parameters correlation to each other. The coefficient of correlations was calculated in the whole wood (sapwood and heartwood together), sapwood, and heartwood parts only.

RESULTS AND DISCUSSION

Extractive content

The extraction yields of each clone using solvents with increasing polarity (n-hexane, methanol, hot water) successively are shown in Figure 1, Figure 2, Figure 3. In general, clone 16 had the highest range of n-hexane-soluble extractive (HEC) and methanol-soluble extractive (MEC) but the lowest value for hot-water-soluble extractive (AEC). Meanwhile, clone 25 had the lowest value of HEC and MEC but the highest level of AEC. The ANOVA found significant differences for both clone (p = <0,01) and radial direction (p = 0,02) of the trees in HEC and in the interaction between clone and radial direction in MEC (p = <0,01) and AEC (p = <0,01) levels. Therefore, Duncan's test was conducted to evaluate the effect of these factors.

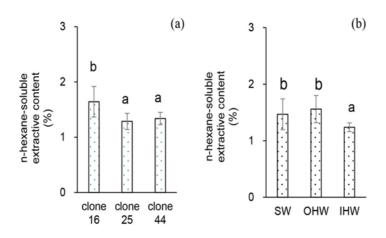


Figure 1: (a) Extractive content of n-hexane (% based O.D. wood) from each clone and (b) three radial directions (SW = sapwood; OHW = outer heartwood; and IHW = inner heartwood) of *Acacia* hybrid (average from three replication). The same letters on the histogram implied no significant differences (p < 0.05 by Duncan's test).

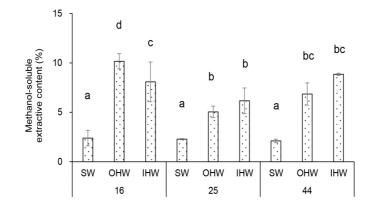


Figure 2: Extractive content of methanol (% based on O.D. wood) from each clone (16, 25, and 44) and radial direction (SW = sapwood; OHW = outer heartwood; IHW = inner heartwood) of *Acacia* hybrid (average from three replication). The same letters on the histogram implied no significant differences (p < 0.05 by Duncan's test).

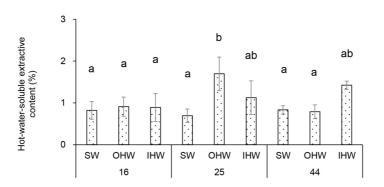


Figure 3: Extractive content of water (% based on O.D. wood) from each clone (16, 25, and 44) and radial direction (SW = sapwood; OHW = outer heartwood; IHW = inner heartwood) of *Acacia* hybrid (average from three replication). The same letters on the histogram implied no significant differences (p < 0.05 by Duncan's test).

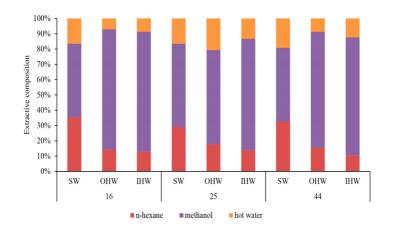


Figure 4: Extractive composition from each clone (16, 25, and 44) and radial direction (SW = sapwood; OHW = outer heartwood; IHW = inner heartwood) of *Acacia* hybrid (average from three replication).

The test showed clone 16 to have the significantly highest level of HEC with 1,64 $\% \pm 0,28$ % while in all clones, the sapwood and outer hardwood part had the highest HEC level with 1,47 $\pm 0,27$ % and 1,56 $\% \pm 0,24$ % respectively as shown in Figure 1a and Figure 1b. Moreover, the outer heartwood of clone 16 had the highest level of MEC with 10,17 $\% \pm 0,78$ % as shown in Figure 2 as well as the AEC amount of the outer heartwood part of clone 25 with 1,70 $\% \pm 0,40$ % as shown in Figure 3. The significant difference between clones indicates the extractive contents were highly affected by the genetic factor of each clone parent's provenance and family.

In comparison with the *A. mangium* from the previous study conducted by Pinto *et al.* (2005), clone 16 had higher lipophilic extractive content, while clone 25 and 44 had a similar range. Meanwhile, the hydrophilic extractive content of all clones is generally similar to *A. auriculiformis* from the previous study (Barry *et al.* 2005). In general, wood with high extractive content, especially lipophilic extractive, is less preferable for pulp and paper material to avoid problems related to pitch deposit and yield reduction (Gutiérrez *et al.* 2004, McLean *et al.* 2014). The high lipophilic and total extractive content of clone 16 has the ability to make it less effective for pulp and paper material compared to the others. However, its high hydrophilic extractive content might indicates high bioactivity of the extract.

Furthermore, the extractive composition of each solvent showed MEC to be dominant in each clone with

a value around 48 % - 79 % based on extract weight as shown in Figure 4. In general, HEC and AEC compositions were highest in sapwood but the decreased in the heartwood. Moreover, MEC concentration was the lowest in sapwood but increased in the heartwood. Theoretically, the formation of heartwood is marked by the death of sapwood cells and the formation of secondary metabolites like phenols (Shmulsky and Jones 2011). The decreased levels of HEC and AEC also indicate the decrease of primary metabolites in the sapwood to heartwood such as starch, fats, and sugar (Taylor *et al.* 2002). A different pattern was observed in the heartwood extractive content for each clone. Clone 16 had a higher extractive content in the outer heartwood part while the others had it in the inner heartwood part. However, the higher concentration in the outer heartwood of clone 16 presumptuously indicates its maturity, as mature woods were known to have higher extractive content (Dünisch *et al.* 2010). The previous study conducted by Nugroho *et al.* (2012) also found a difference in the border of juvenile and mature wood in *A. mangium* from different provenances and there is also a possibility that each clone has a difference in the maturity of the wood.

Total phenolic, flavonoid, and flavanol content

The total phenolic, flavonoid, and flavanol content (TPC, TFC, and TVC) of extracts from *Acacia* hybrid clones are shown in Table 2 and the highest ranges were generally exhibited by methanol-soluble extracts. Moreover, the ANOVA found significant differences in the interaction between clone and radial direction in the TPC of all solvents (p = 0.02; 0.03; and <0.01 respectively). In TFC, those for n-hexane (p = <0.01) and methanol-soluble extracts (p = 0.02; 0.02) significantly affected by the interaction of clone and radial direction factor while those for the water-soluble extract was significantly affected by clone (p = <0.01) and radial direction (p = <0.01) separately. In TVC, n- hexane-soluble extract only affected by radial direction (p = <0.02), while methanol and hot-water-soluble-extract affected by clone (p = 0.03) and radial direction (p = <0.01) separately.

					extracts	5.				
	Radial direction	Total phenolic content			Total flavonoid content			Total flavanol content		
Clone		n-hexane	methanol	hot water	n-hexane	methanol	hot water	n-hexane	methanol	hot water
16	SW	$3,77 \pm$	$117,11 \pm$	$43,92 \pm$	$41,52 \pm$	$29,55 \pm$	$16,18 \pm$	$36,03 \pm$	83,39±	$7,84 \pm$
10	SW	1,11 ^a	$46,22^{a}$	4,19 ^a	2,87 ^g	3,03 ^{ab}	6,21 ^{ab}	$16,02^{ab}$	34,92 ^a	1,92 ^a
	OHW	4,12 ±	$386,56 \pm$	$138,22 \pm$	$35,92 \pm$	$127,48 \pm$	$21,77 \pm$	$38,66 \pm$	$200,53 \pm$	$18,71 \pm$
	Onw	0,46 ^a	27,58 ^{cd}	9,04°	$2,40^{fg}$	14,19 ^{de}	4,46 ^{bc}	8,14 ^{ab}	10,71 ^{bc}	9,43 ^b
	IHW	$3,68 \pm$	$358,46 \pm$	$158,61 \pm$	$15,23 \pm$	$133,71 \pm$	$25,08 \pm$	$45,03 \pm$	$218,21 \pm$	$22,34 \pm$
	ITTW	0,37 ^a	86,80 ^{cd}	31,26°	1,48 ^{bc}	19,83 ^e	2,61 ^{bc}	12,64 ^b	25,14 ^c	$0,78^{b}$
25	25 SW	$5,67 \pm$	$177,60 \pm$	$43,28 \pm$	$23,49 \pm$	$22,07 \pm$	$7,70 \pm$	$56,69 \pm$	$90,10 \pm$	$7,08 \pm$
23		1,95 ^{ab}	20,95 ^{ab}	14,07 ^a	3,80 ^{de}	2,84 ^a	1,92 ^a	13,38 ^{bc}	2,84 ^a	3,77 ^a
	OHW	4,24 ±	$273,80 \pm$	$86,73 \pm$	$29,34 \pm$	$110,18 \pm$	$16,35 \pm$	$28,74 \pm$	$135,17 \pm$	$17,76 \pm$
	OIIW	2,18 ^a	118,29 ^{bc}	41,46 ^b	7,71 ^{ef}	12,64 ^{cd}	6,77 ^{ab}	6,91 ^a	21,37 ^{ab}	3,93 ^b
	IHW	8,29 ±	$440,49 \pm$	$154,77 \pm$	$4,23 \pm$	$132,66 \pm$	$23,70 \pm$	$59,11 \pm$	$169,97 \pm$	25,35 ±
	III W	1,29 ^{bc}	58,87 ^d	29,94°	$0,72^{a}$	3,17 ^e	6,39 ^{bc}	14,65°	3,92 ^{bc}	4,13 ^b
44	SW	$4,18 \pm$	$76,83 \pm$	$58,66 \pm$	$21,73 \pm$	41,15 ±	$19,59 \pm$	$40,89 \pm$	$165,69 \pm$	21,42 ±
	5 10	0,22 ^a	22,06 ^a	10,51 ^{ab}	3,08 ^{cd}	1,92 ^b	6,77 ^{bc}	9,38 ^{ab}	49,19 ^{bc}	5,30 ^b
	OHW	5,49 ±	$381,32 \pm$	$198,92 \pm$	$9,78 \pm$	$101,51 \pm$	$29,37 \pm$	$31,19 \pm$	$179,17 \pm$	21,42 ±
		1,92 ^{ab}	54,78 ^{cd}	20,25 ^d	4,94 ^{ab}	6,77°	9,30°	9,31 ^a	30,63 ^{bc}	4,54 ^b
	IHW	$10,41 \pm$	448,35 ±	$136,65 \pm$	32,19 ±	$113,21 \pm$	$28,67 \pm$	$42,58 \pm$	$188,42 \pm$	23,99 ±
		2,90°	43,95 ^d	7,95°	5,39 ^f	6,39 ^{cd}	1,18 ^c	0,95 ^{ab}	73,02 ^{bc}	3,08 ^b

 Table 2: Total phenolic, flavonoid, and flavanol content of three radial parts from clone 16, 25, and 44

 extracts

Values are expressed as mean \pm SE of three tree each clone.

SW = sapwood; OHW = outer heartwood; IHW = inner heartwood

Equal superscripted letters indicate no significant difference between treatments (p < 0.05) in same solvent or column.

Duncan's test was conducted to evaluate the effect of clone and radial factors on TPC, TFC, and TVC content in each solvent as shown in Table 2. The TPC value was found to be significantly highest in the inner heartwood part of clone 25 with 440,49 mg GAE/g \pm 58,87 mg GAE/g and clone 44 with 448,35 mg GAE/g \pm 43,95 mg GAE/g. Similarly, the highest TFC was found significantly in the inner heartwood part of clone 16 with 133,71 mg GAE/g \pm 19,83 mg QE/g and 25 with 132,66 mg GAE/g \pm 3,17 mg QE/g. Meanwhile, the highest value of TVC was found in the inner heartwood part of clone 16 alone with 218,21 mg GAE/g \pm 25,14 mg CE/g.

The total phenolic content of *A. auriculiformis* has been reported to be five times higher than *A. mangium* (Barry *et al.* 2005) and flavonoids and proanthocyanidins (condensed tannin) are found to be the most dominant phenolic compound founds in *Acacia* heartwood extractive (Foo 1984, Barry *et al.* 2005). The higher phenolic content of *A. auriculiformis* was suspected to be contributing to its better resistance against heart rot and termites (Barry *et al.* 2005) mainly due to the monomeric flavonoids, specifically, 3,4',7,8-tet-ra-hydroxy flavanone and teracacidin, found in greater amount in *A. auriculiformis* (Drewes and Roux 1966, Pietarinen *et al.* 2004, Barry *et al.* 2005). In general, the TPC value of *Acacia* hybrid clones was intermediate between its two parent species and more than *A. mangium*. Moreover, the quantity of flavanol in *A. auriculiformis*, *Acacia* hybrid clones' TVC is 22 times and 6 times higher respectively. This high concentration of flavanol could be caused by the monomer of profisetinindin, a condensed tannin with flavanol monomer identified in *Acacia* species (Seigler 2003, Barry *et al.* 2005).

Correlation between extractive and phenolic contents

Pearson's correlations between each parameter in sapwood and heartwood are shown in Table 3 and in the sapwood, the highest correlation was found between TFC and TVC levels of the methanol-soluble extracts with 0,77*. Further, a strong correlation was found between TPC and TVC in the hot-water-soluble extract with 0,72* and strong negative correlation was found between the TPC and TFC with -0,69*. In the heartwood, the highest negative correlation was found between the extractive content and TPC value of the hot-water-soluble extract with -0,69*. The other strong positive correlations were found between extractive content and TFC level of the n-hexane-soluble extracts with 0,51*, extractive content and TVC level of the methanol-soluble extracts with 0,59* and between TFC and TVC levels of the hot-water-soluble extracts with 0,59*.

Several positive and negative correlations were found in this study. A positive correlation might indicate a high presence of a certain type of compound in the phenol group while a negative one might indicate a low presence. The correlations found in the methanol-soluble extractive of sapwood indicate a high presence of flavanol type compound in the flavonoid group although a low presence of flavonoid was found in the sapwood. In the heartwood part, the result suggested a tree part with higher methanol-soluble extractive content might also have a higher flavanol content. Furthermore, the presence of phenolic compounds in the n-hexane and water-soluble extract were observed not to be dominant.

	Pearson's correlation								
Phenolic		Sapwood		Heartwood					
Phenolic	Extractive content	ТРСН	TFCH	Extractive content	ТРСН	TFCH			
TPCH	-0,39			-0,56*					
TFCH	0,53	-0,45		0,51*	-0,02				
TVCH	-0,14	0,35	-0,50	-0,28	0,25	-0,35			
Phenolic	Extractive content	TPCM	TFCM	Extractive content	TPCM	TFCM			
TPCM	0,50			0,38					
TFCM	-0,02	-0,69*		-0,18	-0,09				
TVCM	0,17	-0,29	0,77*	0,59*	-0,31	0,33			
Phenolic	Extractive content	TPCA		Extractive content	TPCA	TFCA			
TPCA	-0,12			-0,69*					
TFCA	0,47	0,60		-0,26	0,40				
TVCA	0,22	0,72*	0,47	-0,07	0,14	0,49*			

Table 3: Pearson's correlation between extractive content and phenolics content.

TPC = total phenolic content; TFC = total flavonoid content; TVC = total flavonoid content; H = n-hexane; M = methanol; A = water; * marked significant correlation in 0,05 level.

The correlation between extractive and phenolic contents has also been found by Kadir and Hale (2017) in several Malaysian plant species, as total phenol increased in parallel with extractive content. Most phenolic compounds are polar due to the highly electronegative oxygen in its hydroxyl group (Rappoport 2003). Therefore, a low presence of phenolic compounds in the n-hexane-soluble extract was expected indicated by the negative correlation found in the phenol. Meanwhile, a positive correlation found between flavanol and extractive content of sapwood methanol-soluble extract is suspected to be the monomer of profisetinindin, which is a flavanol type phenol identified in previous studies on various *Acacia* species (Seigler 2003, Barry *et al.* 2005).

Antioxidant activity

The antioxidant activities, expressed as IC_{50} (inhibitory concentration (µg/ml) to scavenge the DPPH radical by 50%), are shown in Figure 5 to have only be conducted in the methanol extracts due to its high total phenolic content. The average IC_{50} values of inner heartwood, outer heartwood, and sapwood were found to be 255,77 µg/ml; 289,94 µg/ml, and 680,55 µg/ml respectively. The ANOVA only found a significant difference in the radial direction of *Acacia* hybrid clones (p = <0,01), while Duncan's test was conducted to evaluate the significance in radial factor as shown in Figure 5. The result showed inner heartwood and outer heartwood have significantly similar strongest antioxidant activity and they are weaker than both of the positive controls of gallic acid at 39,00 µg/ml and catechin at 45,52 µg/ml. The IC_{50} value of outer heartwood and inner heartwood were around 7,43 and 6,56 times of gallic acid respectively, and 6,37 and 5,62 times of catechin respectively.

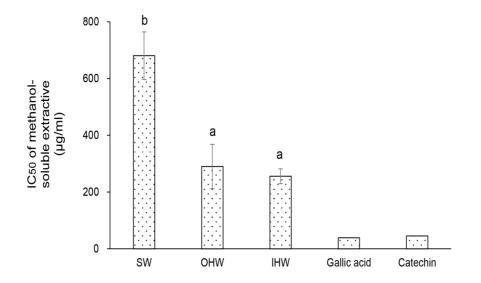


Figure 5: Inhibitory concentration to reduce DPPH radical by 50 % (IC₅₀) in radial direction (SW = sapwood; OHW = outer heartwood; IHW = inner heartwood) of *Acacia* hybrid (average from three replication). The same letters on the histogram implied no significant differences (p = <0,05 by Duncan's test).

Compared to a previous study of *A. mangium* and *A. auriculiformis* methanol extract conducted by Mihara *et al.* (2005), *Acacia* hybrid clones are in between both parent species, where *A. mangium* extract has IC_{50} value 14,53 times of the control gallic acid, and *A. auriculiformis* has 3,76 times. Mihara *et al.* (2005) also found that the antioxidant activity in both species correlated with antifungal activity and laccase enzyme inhibitory of the extract. Moreover, the phenolic compounds were suspected to have the ability to scavenge the hydroxyl radical produced by the laccase enzyme of heart rot fungi. Therefore, this finding shows *Acacia* hybrid clones has a better pest and disease resistance compared to *A. mangium*. Specifically, higher resistance to heart rot is expected from clone 16 and 25 as a great amount of flavonoid was detected in their inner heartwood. Furthermore, the polar extract might also be utilized as a natural source of antioxidant activity than the fractionated and isolated ones, therefore, a stronger antioxidant activity could be achieved with further fractionation and isolation of *Acacia* hybrid extract.

Correlation between phenolics content and antioxidant activity

Total phenolic, flavonoid, and flavanol contents were correlated with the radical scavenging ability to estimate which compounds have more effect on the antioxidant activity. This was measured using Pearson's correlation analysis in the whole radial section, sapwood only, and heartwood only and the coefficients are as presented in Table 4. The highest correlation was found between the antioxidant activity and total flavonoid content with -0,89* as shown in Figure 6a as well as with total phenolic and flavanol content at -0,87* and -0,66* respectively. In the heartwood parts only, a significant correlation was found only in the total phenolic content with -0,75* as presented in Figure 5b and none were found between any phenolic content and the antioxidant activity of the sapwood parts.

	Whole wood			Heartwood			Sapwood		
Part	TPC	TFC	TVC	TPC	TFC	TVC	TPC	TFC	TVC
DPPH IC ₅₀	-0,87*	-0,89*	-0,66*	-0,75*	-0,18	-0,44	0,23	-0,32	-0,16

Table 4: Pearson's correlation between phenolics and antioxidant activity.

^{*} marked significant correlation in 0,01 level. TPC = total phenolic content; TFC = total flavonoid content; TVC = total flavonoi content.

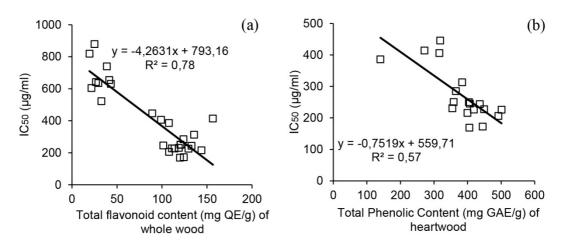


Figure 6: Correlation of (a) total flavonoid content and (b) total phenolic content to IC₅₀ value of DPPH scavenging activity in whole wood and heartwood of *Acacia* hybrid clones wood extract.

Phenolic compounds are known to be able to neutralize the chain-carrying ROO* radicals by transferring a hydrogen atom from their hydroxyl groups (Foti 2007). Previous studies have also found a correlation between phenolic and antioxidant activity levels (Franco *et al.* 2008, Wissam *et al.* 2012, Hennia *et al.* 2018). In *Acacia* species, the compounds 3,4',7,8-tetrahydroxyflavanone and teracacidin are the most abundant flavonoid compound found in *A. mangium* and *A. auriculiformis* to be showing high antioxidant and antifungal activity (Mihara *et al.* 2005). In this research, a significant correlation in heartwood was only found with total phenolic content and this shows there is a possibility some different forms of phenolic compounds other than flavonoids were responsible for the antioxidant activity of the *Acacia* hybrid.

CONCLUSIONS

Clone 16 was determined to have the highest extractive and phenolic contents while clone 25 had the lowest. In comparison with previous studies, the extractive content, total phenolic content, total flavonoid content, and antioxidant activity of *Acacia* hybrid wood extracts intermediate between *A. mangium* and *A. auriculiformis*, while the total flavanol content of the hybrid was much higher.

Clones 25 and 44 could be effective as pulp and paper material to avoid pitch problem and maximize yield because of its lower extractive content. Furthermore, the methanol-soluble extracts of *Acacia* hybrid clones have a good radical scavenging activity and may be used as a potential source of natural antioxidant. From the results of this study, it is assumed that the *Acacia* hybrid clones, especially clone 16, may have potential against heart rot disease compared to *A. mangium*, but not as good as *A. auriculiformis*. However, it would be of interest to further investigate the antifungal activity of *Acacia* hybrid especially against heart rot as well as analyze the phenolic compounds through fractionation, isolation, and antioxidant activity assays using other radical types.

REFERENCES

Baba, S.A.; Malik, S.A. 2015. Determination of total phenolic and flavonoid content, antimicrobial and antioxidant activity of a root extract of *Arisaema jacquemontii* Blume. *J Taibah Univ Sci* 9(4): 449-454. https://doi.org/10.1016/j.jtusci.2014.11.001

Barry, K.M.; Mihara, R.; Davies, N.W.; Mitsunaga, T.; Mohammed, C.L. 2005. Polyphenol in Acacia mangium and Acacia auriculiformis heartwood with reference to heart rot susceptibility. J Wood Sci 51(6): 615-621. https://doi.org/10.1007/s10086-005-0707-x

Coetzee, M.P.A.; Wingfield, B.D.; Golani, G.D.; Tjahjono, B.; Gafur, A.; Wingfield, M.J. 2011. A single dominant Ganoderma species is responsible for root rot of *Acacia mangium* and *Eucalyptus* in Sumatra. *South For* 73(3-4): 175-180. https://doi.org/10.2989/20702620.2011.639488

Diouf, P.N.; Stevanovic, T.; Cloutier, A. 2009. Antioxidant properties and polyphenol contents of trembling aspen bark extracts. *Wood Sci Technol* 43(5-6): 457-470. https://doi.org/10.1007/s00226-009-0240-y

Drewes, S.E.; Roux, D.G. 1966. A new flavan-3,4-diol from *Acacia auriculiformis* by paper ionophoresis. *Biochem J* 98(2): 493-500.

Dünisch, O.; Richter, H.G.; Koch, G. 2010. Wood properties of juvenile and mature heartwood in *Robinia pseudoacacia* L. *Wood Sci Technol* 44(2): 301-313. https://doi.org/10.1007/s00226-009-0275-0

Foo, L.Y. 1984. Condensed tannins: co-occurrence of procyanidins, prodelphinidins and profisetinidins in the heartwood of *Acacia baileyana*. *Phytochemistry* 23(12): 2915-2918. https://doi.org/10.1016/0031-9422(84)83041-1

Foti, M.C. 2007. Antioxidant properties of phenols. J Pharm Pharmacol 59(12): 1673-1685. https://doi.org/10.1211/jpp.59.12.0010

Franco, D.; Sineiro, J.; Rubilar, M.; Sanchez, M.; Jerez, M.; Pinelo, M.; Costoya, N.; Nunez, M.J. 2008. Polyphenols from plant materials: extraction and antioxidant power. *EJEAFChe* 7(8): 3210-3216.

Gutiérrez, A.; del Río, J.C.; Martínez, A.T. 2004. Chemical analysis and biological removal of wood lipids forming pitch deposits in paper pulp manufacturing. In *Environmental Microbiology. Methods in Biotechnology*. Vol. 16. Walker, J.M.; Spencer, J.F.T. (Eds.). Ragout de Spencer A.L. Humana Press: New Jersey, USA.

Hennia, A.; Miguel, M.G.; Nemmiche, S. 2018. Antioxidant activity of *Myrtus communis* L. and *Myrtus nivellei* Batt. & Trab. extracts: a brief review. *Medicines* 5(3): 89. https://doi.org/10.3390/medicines5030089

Kadir, R.; Hale, M.D. 2017. Antioxidant potential and content of phenolic compounds in extracts of twelve selected Malaysian commercial wood species. *Eur J Wood Wood Prod* 75(4): 615-622. https://doi.org/10.1007/s00107-016-1095-1

Kha, L.D. 2000. Studies on natural hybrid of *Acacia mangium* and *A. auriculiformis* in Vietnam. *J Trop* For Sci 12(4): 794-803. https://www.jstor.org/stable/43582411

McLean, D.S.; Stack, R.S.; Richardson, D.E. 2014. The effect of wood extractives composition, pH, and temperature on pitch deposition. *Appita J* 58(1): 52-76. https://search.informit.org/doi/epdf/10.3316/informit.636948896703474

Mihara, R.; Barry, K.M.; Mohammed, C.L.; Mitsunaga, T. 2005. Comparison of anti fungal and antioxidant of *Acacia mangium* and *Acacia auriculiformis*. *J Chem Ecol* 31(4): 789-804. https://doi.org/10.1007/ s10886-005-3544-x

Mohammed, C.L.; Barry, K.M.; Irianto, R.S.B. 2006. Heart rot and root rot in *Acacia mangium* identification and assessment. In *Heart rot and root rot in tropical Acacia plantations*. Potter, K.; Rimbawanto, A.; Beadle, C. (Eds.). ACIAR: Canberra, Australia.

Nugroho, W.D.; Marsoem, S.N.; Yasue, K.; Fujiwara, T.; Nakajima, T.; Hayakawa, M.; Nakaba, S.; Yamagishi, Y.; Jin, H.; Kubo, T.; Funada, R. 2012. Radial variations in the anatomical characteristics and density of the wood of *Acacia mangium* of five different provenances in Indonesia. *J Wood Sci* 58(3): 185-194. https://doi.org/10.1007/s10086-011-1236-4

Pietarinen, S.P.; Willför, S.M.; Sjöholm, R.E.; Holmbom, B.R. 2004. Extractives in *Acacia mangium* and *Acacia crassicarpa* stemwood and knots. In Proceedings of the 58th Appita Conference, 19-21 April 2004, Canberra, Australia.

Pinso, C.; Nasi, R. 1991. The potential use of *Acacia mangium* × *Acacia auriculiformis* hybrid in Sabah. In *Breeding Technologies for Tropical Acacias*. Carron, L.T; Aken, K.M. (Eds.). ACIAR: Canberra, Australia.

Pinto, P.C.; Evtuguin, D.V.; Pascoal, N.C. 2005. Chemical composition and structural features of the macromolecular components of plantation *Acacia mangium* wood. *J Agric Food Chem* 53(20): 7856-7862. https://doi.org/10.1021/jf058081b

Rappoport, Z. 2003. The chemistry of phenols. John Wiley and Sons Ltd: West Sussex, England.

Seigler, D.S. 2003. Phytochemistry of acacia – sensu lato. *Biochem Syst Ecol* 31(8): 845-873. https://doi.org/10.1016/S0305-1978(03)00082-6

Sein, C.C.; Mitlohner, R. 2011. Acacia mangium Willd: Ecology and Silviculture. CIFOR: Bogor, Indonesia. https://www.cifor.org/publications/pdf_files/Books/BCIFOR1105.pdf

Shmulsky, R.; Jones, P.D. 2011. Forest products & wood science – an Introduction. Wiley-Blackwell: Mississippi, USA.

Soon, L.K.; Chiang, L.K. 2012. Influence of different extraction solvents on lipophilic extractives of *Acacia* hybrid in different wood portions. *Asian J Appl Sci* 5(2): 107-116. https://doi.org/10.3923/ajaps.2012.107.116

Sudin, M.; Lee, S.S.; Harun, A.H. 1993. A survey of heart rot in some plantations of *Acacia mangium* in Sabah. *J Trop For Sci* 6(1): 37-47. https://www.jstor.org/stable/43581714

Sunarti, S.; Na'iem, M.; Hardiyanto, E.B.; Indrioko, S. 2013. Breeding strategy of *Acacia* hybrid (*Acacia mangium* x *A. auriculiformis*) to increase plantation productivity in Indonesia. *J Man Hut Trop* 19(2): 128-137. https://doi.org/10.7226/jtfm.19.2.128

Taylor, A.M.; Gartner, B.L.; Morrell, J.J. 2002. Heartwood formation and natural durability: a review. *Wood Fiber Sci* 34(4): 587-611. https://wfs.swst.org/index.php/wfs/article/view/539

Wissam, Z.; Ghada, B.; Wassim, A.; Warid, K. 2012. Effective extraction of polyphenols and proanthocyanidins from pomegranate's peel. *Int J* Pharm Pharm *Sci* 4(3): 675-682.

Yahya, R.; Sugiyama, J.; Silsia, D.; Gril, J. 2010. Some anatomical features of an *Acacia* hybrid, *A. mangium* and *A. auriculiformis* grown in Indonesia with regard to pulp yield and paper strength. *J Trop For Sci* 22(3): 343-351. https://www.jstor.org/stable/23616663